

Effect of hypercholestermic diet on the β - amyloid deposition and microglial cells with some biomarkers alterations in male rats

H.A. Raheem¹ , W. Albazi¹ , R. Altae¹ , T.M. Al-Thuwaini²  and G.H. Jhoni³ 

¹Department of Physiology, Biochemistry and Pharmacology, College of Veterinary Medicine, University of Kerbala, Kerbala, ²Department of Animal Production, College of Agriculture, Al-Qasim Green University, Babel, ³Department of Family Medicine, College of Medicine, University of Kerbala, Kerbala, Iraq

Article information

Article history:

Received 14 April, 2023
Accepted 25 November, 2023
Available online 04 January, 2024

Keywords:

Brain
Cholesterol-raising diet
Lipid profile
Immunohistochemistry

Correspondence:

T.M. Al-Thuwaini
tahreemohammed@gmail.com

Abstract

Unusually high lipid levels define a hypercholesterolemic diet (HCD) and are strongly linked to brain damage and cerebrovascular illnesses. HCD is a chronic brain disorder characterized by cognitive impairment, inflammation, β -amyloid ($A\beta$) deposition, and vascular injury. Recent studies have shown that high cholesterol levels are linked to AD pathology. This investigation aims to check out the physiological modifications of the brain that happen from receiving a high-cholesterol diet. We have previously shown that high brain cholesterol levels promote $A\beta$ accumulation and oxidative stress. The experiment employed sixteen male rats. Which was split into two groups: the oversight group (8 rats) and the cholesterol group (8 rats), the latter of which received a 1% supplement of cholesterol in their food 28 days later, rats' blood was drawn for biochemical analysis and brain tissues were removed and processed for light microscopy inspection using H&E and CD68, an immunohistochemistry marker for microglia cells. Brain samples were homogenized to measurement of the $A\beta$. Significant increase in serum lipid profile, $A\beta$, Acetylcholinesterase (AChE), and Malnodialdehyde (MDA) levels, while Significant decrease of serum HDL-C and serum Glutathione (GSH) levels in cholesterol group compared to the control group. Rats' brains had visible morphological alterations, having pyknotic nuclei of degenerating neurons, lack of neurons, and pathological alteration in morphology of microglial cells in the cholesterol group compared to the control group. We concluded that an HCD has negative biochemical alterations linked to brain anatomical changes.

DOI: [10.33899/ijvs.2023.139742.2973](https://doi.org/10.33899/ijvs.2023.139742.2973), ©Authors, 2023, College of Veterinary Medicine, University of Mosul.
This is an open access article under the CC BY 4.0 license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction

Cholesterol is a waxy, fat-like substance with pivotal pathophysiological relevance (1). A hydrophobic substance known as cholesterol is carried through the circulation by lipoproteins, proteins; John Gofman used an ultracentrifuge to isolate the lipoproteins in plasma (1). Multiple physiological conditions, such as fat, cardiovascular disease, and Alzheimer's disease, are brought on by high cholesterol levels (2). Additionally, hypercholesterolemia, or elevated plasma cholesterol levels, are linked to male

infertility because they cause the male reproductive system's malfunction (2). A high-fat diet can worsen reactive stress and inflammation in the brain and negatively impact cognitive function (3). An environment that is susceptible to harm from a high-fat diet is the aging brain (3). The brain is particularly vulnerable to oxidative stress, which can be produced by an elevated cholesterol diet (4). A high cholesterol level and oxidative stress are substantial risk factors for various illnesses, including dementia and many central nervous system problems (4). Hypercholesterolemia increases levels of β -amyloid ($A\beta$), a

peptide that accumulates in Alzheimer's disease brains (5). For the brain to continue to have a regular shape and function, cholesterol is a crucial substance. However, a long-term high-cholesterol diet can result in several degenerative changes to the brain, including the buildup of β -amyloid ($A\beta$), hyperphosphorylation of Tau, reactive gliosis, neuroinflammation, cell mortality, and synaptic degeneration (6). These abnormal alterations interact with one another in intricate ways, impairing memory and contributing to the development of Alzheimer's disease (AD) (6). Oxysterols, oxidized versions of cholesterol, are more readily transported to the brain by high-fat diets, which may help explain the correlation between blood and brain cholesterol levels. It is thought that these cholesterol compounds may also help cerebral cholesterol carry out its intended tasks in the brain. In light of this, new research suggests that oxysterols are essential signaling molecules for brain processes (7). To avoid abnormal biochemical processes that result from an excessive buildup of cholesterol in tissues, cholesterol homeostasis is carefully kept. Although dyslipidemia impairs endocrine function and impairs reproduction in male rodents, no obvious regulatory variables or processes have been established (2). However, it is unclear what early hypercholesterolemia-related events cause brain degeneration (6).

The current study aims to investigate the effect of the hypercholesterolemic diet on some oxidative stress parameters, beta-amyloid deposition, and microglial cell activity by immunohistochemistry on the brain in male rats.

Materials and methods

Ethical approve

Under the reference number UOK. The VET.PH.2022.046 study was conducted at the Kerbala University, College of Veterinary Medicine's anatomical facility in Iraq.

Experimental protocol

16 white male rats' weight 200 ± 20 g were used in this research and came from the College of Pharmacy at the University of Kerbala in Iraq. They ranged in age from 11-14 weeks, and the animals were housed in clean, specialized plastic enclosures. They receive the proper air and surroundings. We utilized a 12-hour light cycle and a relative humidity of 55%. They were retained for two weeks to adjust to the usual testing conditions. The experiment began on the 15th of December and ended on the 12th of January. The temperature was maintained at 23-26°C using a room thermostat; the room air was changed continuously using a ventilation vacuum, and the animal fed on the pellet of freshly prepared ration.

Experimental design

16 white male rats were arbitrarily split into two groups and given the following treatments. 8 rats of this group were given a regular meal orally as the comparison group. 8 rats received cholesterol. Rats in this group were given a meal rich in cholesterol for 28 days, comprising 1% cholesterol (w/w) (8).

The taking of blood samples

Fasting after 28 days of the trial, blood samples were taken, and before the blood was born, the animals were controlled and made comfortable by inhaling chloroform. Sterile medical syringes of 5 ml were accustomed to extract 5 ml of cardiac blood using the heart puncture method, and the blood was then put into the serum was centrifuged in a unique gel tube that did not contain an anticoagulant at a speed of 4000 revolutions per minute for 5 minutes. Once the serum had been separated, it was stored in Eppendorf tubes and kept in the fridge at -30°C while the assays worked.

Collecting of organs for the histological section

Rats were killed by chloroform anesthesia following the completion of the experiment, and the animals were dissected to extract sample brains. The organs were enumerated and then preserved in 10% formalin in sterile plastic containers until the histological section was carried out.

Gathering of the brain tissues

Weighed brain samples were crushed using Squishers in 200 l of 0.1 N perchloric acid for 4 minutes (to ensure the tissue was evenly distributed). The supernatants were meticulously removed after centrifuging the homogenates for 30 minutes in a cold environment (4°C). The supernatants can be collected immediately for analysis or frozen at -80°C. Take 500 ml of anhydrous glacial acetic acid, 25 ml of acetic anhydride, and about 8.5 ml of perchloric acid (24h). Complete the combination with 1000 cc of anhydrous glacial acetic acid-the beta-amyloid measurement.

An immunohistochemistry

Stain (CD68) was used to detect microglial cells.

Statistical analysis

In the statistical program Graph Pad Prism 8.0, the t-test was used, and $P\leq 0.05$ was chosen as the standard of significance. The data points were shown as mean \pm SD.

Results

Blood serum parameters

Figure 1 in the present study showed a significant increase in the TC, TG, LDL-C, and VLDL-C in the cholesterol group 79.75 ± 3.19 , 78.00 ± 7.90 , 14.41 ± 2.91 , and 31.46 ± 5.12 compare to the control group 46.63 ± 3.46 , 43.75 ± 5.20 , 8.25 ± 1.36 , and 10.97 ± 7.17 respectively. In contrast, figure 1 showed a significant decrease in the HDL-C 10.13 ± 2.94 in the cholesterol group when

compared to the control group 40.88 ± 9.71 and a significant increase in the MDA in the cholesterol group 0.28 ± 0.18 compare with control group 0.07 ± 0.026 . Figure 1 revealed a considerable reduction in the GSH in the cholesterol group 336.91 ± 64.82 compared with the control group 437.8 ± 21.20 , while a significant increase in the AChE 15.74 ± 2.15 in cholesterol group when compared to the control group 9.331 ± 0.38 . A significant increase in the β -Amyloid in the cholesterol group 129.20 ± 7.03 compared with the control group 43.11 ± 9.76 .

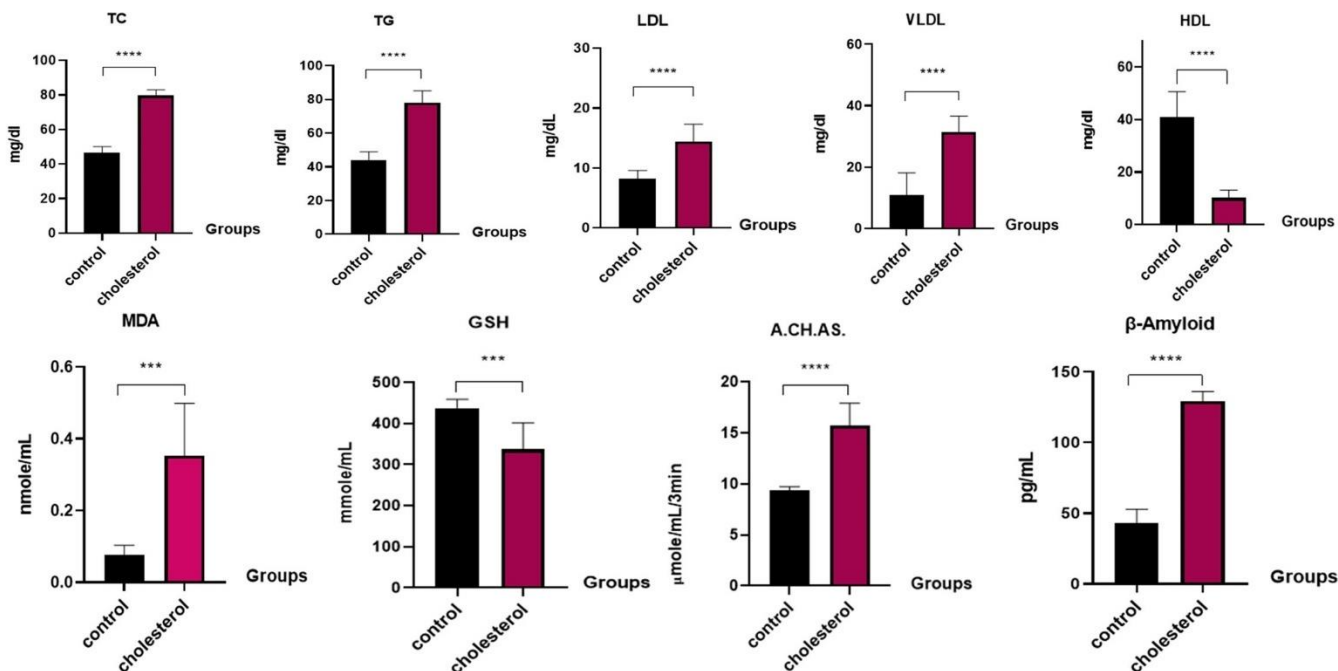


Figure 1: Effect of hypercholesteremic diet on the serum parameters concentration in the male rats.

Histopathological and immunohistochemistry examination

The control rats displayed normal morphology of microglial cells, while the histological analysis of hypercholesterolemia rats revealed a pathological alteration in the morphology of the microglial cells, resulting in a decrease in phagocytic activity and changes in bodily features with slight shortening in length. Additionally, astrocyte degenerative changes and marked pyknotic changes in oligodendrocytes were observed. Immunohistochemistry staining further confirmed the normal and altered morphology of microglia as shown in figure 2.

Discussion

Cholesterol is a widely recognized important lipid membrane modulator structure and fluidity, and as such, it is crucial for maintaining transmembrane transmission

within and between cellular divisions. Cholesterol amounts in membranes are necessary for living. Despite making up only 2.2% of the body weight, the central nervous system (CNS) includes up to 25% of the body's overall cholesterol (9-12).

In the current research, a diet high in cholesterol improved TC, TG, LDL-C, and VLDL-C compared to the control group. At the same time, HDL-C showed a significant decrease in the cholesterol group in compares with the control group. This result is in agreement with (3,4,13-17). Although it is unclear how cholesterol can impact brain processes, its oxysterols and oxygenated products can cross lipophilic membranes even though cholesterol itself cannot (9,18). According to the study's findings, blood GSH levels significantly decreased and increased in the MDA in the cholesterol group compared to the control group. These results are in agreement with (19-23). It also showed a significant increase in the β -amyloid(A β) and AChE, which agrees with (18,24).

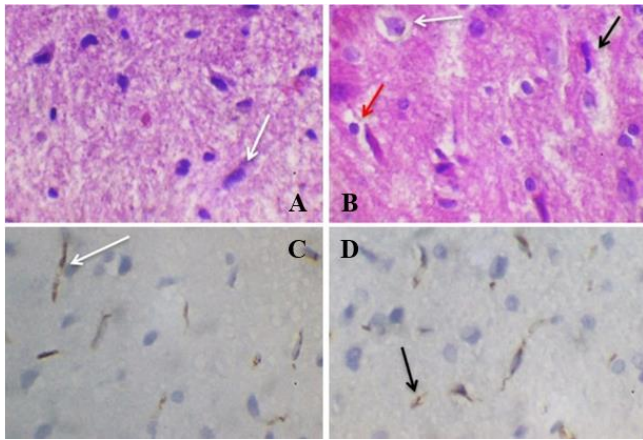


Figure 2: (A) Control rat showing normal morphology of microglial cells (cerebral macrophages) (white arrow) (B) histological picture showing pathological alteration in morphology of the microglial cell, towards lowering its phagocytic activity resulting in changes in its bodily features represented by slight shortening in their length (black arrow), astrocyte degenerative changes (white arrow) and marked pyknotic changes in oligodendrocytes (red arrow). (C) Immunohistochemistry staining for sections of a control rat showing normal morphology of microglial cells (cerebral macrophages) (white arrow) (D) histological picture showing pathological alteration in microglial cell morphology towards lowering its phagocytic activity resulting in changes in bodily features represented by slight shortening in their length (black arrow).

MDA levels significantly increased in hypercholesterolemic rats, and the antioxidant enzyme system was suppressed in the rats' liver, renal, heart, and brain tissues (20-23). In the etiology of many illnesses, oxidative stress is a key player. The relationship between hypercholesterolemia and oxidative stress in producing endothelial dysfunction in brain arterioles was observed despite the lack of atherosclerotic lesions. These were demonstrated and verified in this research by higher MDA concentration (9,25).

The antioxidant protection against A β induced mitochondrial ROS is reduced in neuronal cells with high cholesterol levels due to impaired mitochondrial GSH transfer (19,26). Before the formation of amyloid plaques and cognitive deficits, mitochondrial dysfunction is recognized as a frequent early occurrence in Alzheimer's disease (AD). Intracellular A can alter mitochondrial function and promote the generation of ROS, which is further exacerbated by cholesterol-mediated depletion of mitochondrial glutathione (GSH) (19,27). The pathogenesis of AD, including neuroinflammation, is accelerated and made worse by brain cholesterol enrichment by enhancing the mitochondrial oxidative damage caused by A β (24). Different experimental studies have demonstrated that

elevated cholesterol encourages the synthesis and deposition of A β and that this abnormality has repeatedly been linked to Alzheimer's disease (AD) (25). We have also shown that the impairment of autophagy by intracellular cholesterol accumulation impacts A β elimination (26). Recently, we've discovered that cholesterol

Radicals and compounds called ROS are produced when molecular oxygen undergoes an imperfect reduction. They are created in tiny amounts due to four consecutive one-electron reductions of oxygen that result in the creation of water. They are essential for communication and are required to keep cells' homeostasis (27). A β adequate stimulus for causing the formation of A β is free radicals (28-31). That could have significant effects because it indicates that mitochondrial malfunction plays a crucial role in the etiology of A β despite mitochondrial malfunction and stress from oxidation; because mitochondrial malfunction associated with increased mitochondrial ROS generation is known to be strongly tied to the aging process, this might start a loop of increasing amyloidogenic APP (32).

Elevated acetylcholinesterase (AChE) activity in the hypercholesterolemic rats demonstrated cholinergic dysfunction; this increased AChE activity accelerated the hydrolysis of ACh and resulted in its scarcity at the synaptic connections (19). AChE activity alterations and variations in its variant in the blood, cerebrospinal fluid (CSF), and brain (22,33). There is a rise in acetylcholinesterase (AChE) activity within and around amyloid aggregates (30). AChE levels could rise due to a high rate of neuronal degeneration, which increases the amount of unbound AChE in the body and increases the cytotoxicity of amyloid components (34). There is a strong correlation between the prevalence of amyloid structures and changed glycosylation of some AChE forms in AD (22,35).

The hypercholesterolemic rats showed varying degrees of degenerative changes and Lack of neurons, oligodendrocytes with degenerating pyknotic nuclei, vacuolation of the neuropil linked to the growth of neuroglial cells, and abnormalities in the proliferation, phagocytosis, and clearing of microglia were all present (9). Hyperlipidemia The cerebrovascular system acquired atherosclerotic plaques, which resulted in reduced blood flow to the brain, reactive stress, and inflammation (9).

An essential molecule for maintaining brain balance, cholesterol is primarily produced by oligodendrocytes and astrocytes in the brain (9,11). Although the production of cholesterol by oligodendrocytes for the myelination process is well known, it has been demonstrated that astrocytes also create cholesterol for neural cells (36,37). Cholesterol is needed for active axonal development, synapse formation, and remodeling but cannot be supplied by the neuron's distant cell body (34). Microglia are crucial for maintaining brain balance and could be a therapeutic focus for neuronal injury (38-42). Microglia's phagocytic clearing is essential

for regulating neuronal balance and initiating tissue healing (39-43). The BBB was disrupted by the oxidized lipids, particularly oxysterol, which also caused endothelial failure (9). The BBB's injury and the preexisting cerebral hypoxia caused the brain to go through neurodegenerative processes (9). Due to up-regulated lipid metabolism and phagocytosis genes, microglia could not switch to an active state for A clearance, possibly contributing to the growth in amyloid pathology in AD patients (39,42,44). Damaged microglia with high cholesterol buildup may result from downregulated cholesterol production pathways (35). Growing evidence suggests that the cellular phenotypes and roles of microglia change as diseases and the development of the brain advance and Hyperlipidemia the cerebrovascular system acquired atherosclerotic plaques, which resulted in reduced blood flow to the brain, reactive stress, and inflammation (45,46).

Conclusion

Viewing our outcomes, we discovered that hypercholesterolemic diets are a prevalent danger factor for brain damage. We concluded that hypercholesterolemia has negative biochemical alterations linked to brain anatomical changes.

Acknowledgment

The University of Kerbala's College of Veterinary Medicine has been recognized for facilitating this investigation.

Conflict of interest

There is absolutely no apparent conflict, according to the author.

Reference

1. YDuan Y, Gong K, Xu S, Zhang F, Meng X, Han J. Regulation of cholesterol homeostasis in health and diseases: From mechanisms to targeted therapeutics. *Signal Transduct Target Ther*. 2022;7(1):265. DOI: [10.1038/s41392-022-01125-5](https://doi.org/10.1038/s41392-022-01125-5)
2. Lim W, Bae H, Sohn JY, Jeong W, Kim SH, Song G. Dietary cholesterol affects expression of prostatic acid phosphatase in reproductive organs of male rats. *Biochem Biophys Res Commun*. 2015;456(1):421-7. DOI: [10.1016/j.bbrc.2014.11.100](https://doi.org/10.1016/j.bbrc.2014.11.100)
3. Ledreux A, Wang X, Schultzberg M, Granholm AC, Freeman LR. Detrimental effects of a high fat/high cholesterol diet on memory and hippocampal markers in aged rats. *Behav Brain Res*. 2016;312:294-304. DOI: [10.1016/j.bbr.2016.06.012](https://doi.org/10.1016/j.bbr.2016.06.012)
4. Mokhtarzadeh Bazargani M, Naderi G, Roghani M, Esmaili Jamaat E, Hasheminejad SA. Evaluating the effect of *Trachyspermum ammi* (ajwain) hydro-alcoholic extract on oxidative stress markers and cholinesterase activity in brain of male rats fed by a high cholesterol diet. *Daneshvar Med*. 2021;29(1):59-69. DOI: [10.22070/DANESHMED.2021.12757.0](https://doi.org/10.22070/DANESHMED.2021.12757.0)
5. Sharma S, RP JP, Schommer E, Feist G, Ghribi O. Hypercholesterolemia-induced A β accumulation in rabbit brain is associated with alteration in IGF-1 signaling. *Neurobiol Dis*. 2008;32(3):426-32. DOI: [10.1016/j.nbd.2008.08.002](https://doi.org/10.1016/j.nbd.2008.08.002)
6. Chen YL, Wang LM, Chen Y, Gao JY, Marshall C, Cai ZY, Hu G, Xiao M. Changes in astrocyte functional markers and β -amyloid metabolism-related proteins in the early stages of hypercholesterolemia. *Neurosci*. 2016;316:178-91. DOI: [10.1016/j.neuroscience.2015.12.039](https://doi.org/10.1016/j.neuroscience.2015.12.039)
7. Loera-Valencia R, Goikolea J, Parrado-Fernandez C, Merino-Serrais P, Maioli S. Alterations in cholesterol metabolism as a risk factor for developing Alzheimer's disease: Potential novel targets for treatment. *J Steroid Biochem Mol Biol*. 2019;190:104-14. DOI: [10.1016/j.jsbmb.2019.03.003](https://doi.org/10.1016/j.jsbmb.2019.03.003)
8. Wang Q, Du Z, Zhang H, Zhao L, Sun J, Zheng X, Ren F. Modulation of gut microbiota by polyphenols from adlay (*Coix lacryma-jobi L. var. ma-yuen Stapf.*) in rats fed a high-cholesterol diet. *Int J Food Sci Nutr*. 2015;66(7):783-9. DOI: [10.3109/09637486.2015.1088941](https://doi.org/10.3109/09637486.2015.1088941)
9. Metwally ES, Karawya FS. Neuroprotective effects of purslane seeds against adverse effects induced by experimental hyperlipidemia on frontal cortex and cerebellum in young male albino rats. *Int J Clin Exp Med Sci*. 2015;1:3. DOI: [10.11648/j.ijcems.20150103.14](https://doi.org/10.11648/j.ijcems.20150103.14)
10. Liu JP, Tang Y, Zhou S, Toh BH, McLean C, Li H. Cholesterol involvement in the pathogenesis of neurodegenerative diseases. *Mol Cell Neurosci*. 2010;43(1):33-42. DOI: [10.1016/j.mcn.2009.07.013](https://doi.org/10.1016/j.mcn.2009.07.013)
11. Pfrieger FW. Cholesterol homeostasis and function in neurons of the central nervous system. *Cell Mol Life Sci*. 2003;60:1158-71. DOI: [10.1007/s00018-003-3018-7](https://doi.org/10.1007/s00018-003-3018-7)
12. Karam I, Ma N, Yang YJ, Li JY. Induce hyperlipidemia in rats using high fat diet investigating blood lipid and histopathology. *J Hematol Blood Disord*. 2018;4(1):104. DOI: [10.15744/2455-7641.4.104](https://doi.org/10.15744/2455-7641.4.104)
13. Diao SL, Sun JW, Ma BX, Li XM, Wang D. Influence of crocetin on high-cholesterol diet induced atherosclerosis in rats via anti-oxidant activity together with inhibition of inflammatory response and p38 MAPK signaling pathway. *Saudi J Biol Sci*. 2018;25(3):493-9. DOI: [10.1016/j.sjbs.2016.11.005](https://doi.org/10.1016/j.sjbs.2016.11.005)
14. Binayi F, Moslemi M, Khodaghali F, Hedayati M, Zardooz H. Long-term high-fat diet disrupts lipid metabolism and causes inflammation in adult male rats: Possible intervention of endoplasmic reticulum stress. *Arch Physiol Biochem*. 2023;129(1):204-12. DOI: [10.1080/13813455.2020.1808997](https://doi.org/10.1080/13813455.2020.1808997)
15. Aldabbagh EH, Othman LK, Ismail HK. The effects of ghee administration in comparison to sunflower seeds oil on liver tissue and some biochemical parameters in rats. *Iraqi J Vet Sci*. 2022;36:241-8. DOI: [10.33899/ijvs.2022.136030.2558](https://doi.org/10.33899/ijvs.2022.136030.2558)
16. Abdul-Majeed AF, Rahawi GA, Abdul-Rahman SY. Supplementation of broiler drinking water with zinc sulfate and its impact on physiological performance. *Iraqi J Vet Sci*. 2022;36:131-6. DOI: [10.33899/ijvs.2022.135823.2524](https://doi.org/10.33899/ijvs.2022.135823.2524)
17. Jasim RF, Allwsh TA. Effect of plasma isolated Orexin-A on the regulation of metabolites in male rats. *Iraqi J Vet Sci*. 2021;35(3):451-7. DOI: [10.33899/ijvs.2020.127001.1429](https://doi.org/10.33899/ijvs.2020.127001.1429)
18. Vejux A, Malvitte L, Lizard G. Side effects of oxysterols: cytotoxicity, oxidation, inflammation, and phospholipidosis. *Braz J Med Biol Res*. 2008;41(7):1-12. DOI: [10.1590/s0100-879x2008000700001](https://doi.org/10.1590/s0100-879x2008000700001)
19. de Dios C, Abadin X, Roca-Agujetas V, Jimenez-Martinez M, Morales A, Trullas R, Mari M, Colell A. Inflammasome activation under high cholesterol load triggers a protective microglial phenotype while promoting neuronal pyroptosis. *Transl Neurodegener*. 2023;12(1):1-23. DOI: [10.1186/s40035-023-00343-3](https://doi.org/10.1186/s40035-023-00343-3)
20. Hazzaa SM, Eldaim MA, Fouda AA, Mohamed AS, Soliman MM, Elgizawy EI. Intermittent fasting ameliorated high-fat diet-induced memory impairment in rats via reducing oxidative stress and glial fibrillary acidic protein expression in brain. *Nutrients*. 2020;13(1):10. DOI: [10.3390/nu13010010](https://doi.org/10.3390/nu13010010)
21. Otunola GA, Oloyede OB, Oladiji AT, Afolayan AJ. Selected spices and their combination modulate hypercholesterolemia-induced oxidative stress in experimental rats. *Biol Res*. 2014;47(1):1-6. DOI: [10.1186/0717-6287-47-5](https://doi.org/10.1186/0717-6287-47-5)

22. Mahdi SS, Albazi W, Hussain Al-Aameli M. The Beneficial effect of glutathione in the protection of the central nerves system from damage induced by D-galactose. Proc 1st Int Ninevah Conf Med Sci. 2021;70-74. DOI: [10.2991/ahsr.k.211012.012](https://doi.org/10.2991/ahsr.k.211012.012)
23. Al-Safo AA, AlDulaimi LH. Effect of orlistat and aquatic extract of *Rosmarinus officinalis* leaves in histopathological changes in kidney of albino rat. Iraqi J Vet Sci. 2022;36(2):393-400. DOI: [10.33899/ijvs.2021.130400.1813](https://doi.org/10.33899/ijvs.2021.130400.1813)
24. Fouad GI. Combination of omega 3 and coenzyme Q10 exerts neuroprotective potential against hypercholesterolemia-induced Alzheimer's-Like disease in rats. Neurochem Res. 2020;45(5):1142-55. DOI: [10.1007/s11064-020-02996-2](https://doi.org/10.1007/s11064-020-02996-2)
25. Greash ZA, Abbas OA, El-Sayyad HI. Role of drenched barley in improving cerebral structure and function in hypercholesterolemic albino rat dam. Alfarama J Basic Appl Sci. 2021;2(1):70-80. DOI: [10.21608/ajbas.2020.37472.1026](https://doi.org/10.21608/ajbas.2020.37472.1026)
26. Roca-Agüjetas V, Barbero-Camps E, de Dios C, Podlesniy P, Abadin X, Morales A, Marí M, Trullàs R, Colell A. Cholesterol alters mitophagy by impairing optineurin recruitment and lysosomal clearance in Alzheimer's disease. Mol Neurodegener. 2021;16(1):1-26. DOI: [10.1186/s13024-021-00435-6](https://doi.org/10.1186/s13024-021-00435-6)
27. Swerdlow RH. Mitochondria and mitochondrial cascades in Alzheimer's disease. J Alzheimer's Dis. 2018;62(3):1403-16. DOI: [10.3233/JAD-170585](https://doi.org/10.3233/JAD-170585)
28. Barbero-Camps E, Fernández A, Martínez L, Fernández-Checa JC, Colell A. APP/PS1 mice overexpressing SREBP-2 exhibit combined A β accumulation and tau pathology underlying Alzheimer's disease. Hum Mol Genet. 2013;22(17):3460-76. DOI: [10.1093/hmg/ddt201](https://doi.org/10.1093/hmg/ddt201)
29. Wang H, Kulas JA, Wang C, Holtzman DM, Ferris HA, Hansen SB. Regulation of beta-amyloid production in neurons by astrocyte-derived cholesterol. Proc Natl Acad Sci. 2021;118(33):e2102191118. DOI: [10.1073/pnas.2102191118](https://doi.org/10.1073/pnas.2102191118)
30. Barbero-Camps E, Roca-Agüjetas V, Bartolessis I, de Dios C, Fernández-Checa JC, Marí M, Morales A, Hartmann T, Colell A. Cholesterol impairs autophagy-mediated clearance of amyloid beta while promoting its secretion. Autophagy. 2018;14(7):1129-54. DOI: [10.1080/15548627.2018.1438807](https://doi.org/10.1080/15548627.2018.1438807)
31. Devasagayam TP, Tilak JC, Boloor KK, Sane KS, Ghaskadbi SS, Lele RD. Free radicals and antioxidants in human health: Current status and future prospects. J Assoc Physicians India. 2004;52(794804):4. [available at www.japi.org](http://www.japi.org)
32. Leuner K, Schütt T, Kurz C, Eckert SH, Schiller C, Occhipinti A, Mai S, Jendrach M, Eckert GP, Kruse SE, Palmeter RD. Mitochondrion-derived reactive oxygen species lead to enhanced amyloid beta formation. Antioxid Redox Signal. 2012;16(12):1421-33. DOI: [10.1089/ars.2011.4173](https://doi.org/10.1089/ars.2011.4173)
33. García-Ayllón MS, Silveyra MX, Sáez-Valero J. Association between acetylcholinesterase and β -amyloid peptide in Alzheimer's cerebrospinal fluid. Chem Biol Interact. 2008;175(1-3):209-15. DOI: [10.1016/j.cbi.2008.04.047](https://doi.org/10.1016/j.cbi.2008.04.047)
34. Zhang XJ, Greenberg DS. Acetylcholinesterase involvement in apoptosis. Front Mol Neurosci. 2012;5:40. DOI: [10.3389/fnmol.2012.00040](https://doi.org/10.3389/fnmol.2012.00040)
35. Rahman MM, Lendel C. Extracellular protein components of amyloid plaques and their roles in Alzheimer's disease pathology. Mol Neurodegener. 2021;16(1):1-30. DOI: [10.1186/s13024-021-00465-0](https://doi.org/10.1186/s13024-021-00465-0)
36. Hayashi H, Campenot RB, Vance DE, Vance JE. Glial lipoproteins stimulate axon growth of central nervous system neurons in compartmented cultures. J Biol Chem. 2004;279(14):14009-15. DOI: [10.1074/jbc.M313828200](https://doi.org/10.1074/jbc.M313828200)
37. Wahrle SE, Jiang H, Parsadanian M, Legleiter J, Han X, Fryer JD, Kowalewski T, Holtzman DM. ABCA1 is required for normal central nervous system ApoE levels and for lipidation of astrocyte-secreted ApoE. J Biol Chem. 2004;279(39):40987-93. DOI: [10.1074/jbc.M407963200](https://doi.org/10.1074/jbc.M407963200)
38. Hering H, Lin CC, Sheng M. Lipid rafts in the maintenance of synapses, dendritic spines, and surface AMPA receptor stability. J Neurosci. 2003;23(8):3262-71. DOI: [10.1523/jneurosci.23-08-03262.2003](https://doi.org/10.1523/jneurosci.23-08-03262.2003)
39. Wang N, Wang M, Jeevaratnam S, Rosenberg C, Ikezu TC, Shue F, Doss SV, Alnobani A, Martens YA, Wren M, Asmann YW. Opposing effects of apoE2 and apoE4 on microglial activation and lipid metabolism in response to demyelination. Mol Neurodegener. 2022;17(1):1-20. DOI: [10.1186/s13024-022-00577-1](https://doi.org/10.1186/s13024-022-00577-1)
40. Krasemann S, Madore C, Cialic R, Baufeld C, Calcagno N, El Fatimy R, Beckers L, O'loughlin E, Xu Y, Fanek Z, Greco DJ. The TREM2-APOE pathway drives the transcriptional phenotype of dysfunctional microglia in neurodegenerative diseases. Immunity. 2017;47(3):566-81. DOI: [10.1016/j.immuni.2017.08.008](https://doi.org/10.1016/j.immuni.2017.08.008)
41. Magno L, Bunney TD, Mead E, Svensson F, Bictash MN. TREM2/PLC γ 2 signalling in immune cells: Function, structural insight, and potential therapeutic modulation. Mol Neurodegener. 2021;16(1):1-6. DOI: [10.1186/s13024-021-00436-5](https://doi.org/10.1186/s13024-021-00436-5)
42. Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Sternfeld R, Ulland TK, David E, Baruch K, Lara-Astaiso D, Toth B, Itzkovitz S. A unique microglia type associated with restricting development of Alzheimer's disease. Cell. 2017;169(7):1276-90. DOI: [10.1016/j.cell.2017.05.018](https://doi.org/10.1016/j.cell.2017.05.018)
43. Neumann H, Kotter MR, Franklin RJ. Debris clearance by microglia: an essential link between degeneration and regeneration. Brain. 2009;132(2):288-95. DOI: [10.1093/brain/awn109](https://doi.org/10.1093/brain/awn109)
44. Whalley K. A protective population?. Nat Rev Neurosci. 2017;18(8):454. DOI: [10.1038/nrn.2017.83](https://doi.org/10.1038/nrn.2017.83)
45. Chausse B, Kakimoto PA, Kann O. Microglia and lipids: how metabolism controls brain innate immunity. Semin Cell Dev Biol. 2021;112:137-144. DOI: [10.1016/j.semcdb.2020.08.001](https://doi.org/10.1016/j.semcdb.2020.08.001)
46. Marschallinger J, Iram T, Zardeneta M, Lee SE, Lehallier B, Haney MS, Pluvillage JV, Mathur V, Hahn O, Morgens DW, Kim J. Lipid-droplet-accumulating microglia represent a dysfunctional and proinflammatory state in the aging brain. Nat Neurosci. 2020;23(2):194-208. DOI: [10.1038/s41593-019-0566-1](https://doi.org/10.1038/s41593-019-0566-1)

تأثير اتباع نظام غذائي مفرط الكوليسترول على ترسب بيتا اميلويد والخلايا الدبقية مع بعض التغيرات في المؤشرات الحيوية في ذكور الجرذان

حسين عدنان رحيم¹، وفاق البازي¹، راند العظية¹، تحرير محمد الثويني² و غصون هادي جوني³

¹ فرع الفلسفة والكيمياء والحياتية والأدوية، كلية الطب البيطري، جامعة كربلاء، كربلاء، أقسم الإنتاج الحيواني، كلية الزراعة، جامعة القاسم الخضراء، بابل، ² فرع طب الأسرة، كلية الطب، جامعة كربلاء، كربلاء، العراق

الخلاصة

يتميز النظام الغذائي بفرط كوليسترول الدم بارتفاع مستويات الدهون بشكل غير طبيعي ويرتبط بشكل إيجابي بأمراض الأوعية الدموية الدماغية وتلف الدماغ. النظام الغذائي بفرط كوليسترول هو اضطراب مزمن في الدماغ يتميز بضعف الإدراك والالتهاب وترسب البيتا أميلويد وتلف الأوعية الدموية. أظهرت الدراسات الحديثة أن ارتفاع مستويات الكوليسترول في الدم يرتبط بأمراض الزهايمر. الهدف من هذا البحث هو دراسة التغيرات الهيكلية التي تحدث في الدماغ بعد اتباع نظام غذائي عالي الكوليسترول. لقد أظهرنا سابقاً أن ارتفاع مستويات الكوليسترول في الدماغ يعزز تراكم البيتا أميلويد والإجهاد التأكسدي، وقد تم استخدام ستة عشر من ذكورا الجرذان في الدراسة، والمواد والطرق، والتي تم تقسيمها إلى مجموعتين؛

انخفاض كبير في مستويات المصل (بروتين دهني عالي الكثافة) ومصل الكلوتاثيون في مجموعة الكوليسترول مقارنة بمجموعة التحكم. كانت التغيرات الهيكلية المصاحبة في دماغ الجرذان واضحة مثل النوى التقرحية للخلايا العصبية المتدهورة، ونقص الخلايا العصبية والتغيرات المرضية في مورفولوجيا الخلايا الدبقية الصغيرة في مجموعة الكوليسترول مقارنة بمجموعة التحكم. نستخلص إلى أن النظام الغذائي لفرط كوليسترول الدم له تغيرات كيميائية حيوية ضارة مرتبطة بالتغيير الهيكلي في الدماغ.

المجموعة الضابطة (٨ الجرذان) مجموعة الكوليسترول (٨ الجرذان) الذين تم إطعامهم النظام الغذائي المكمل بالكوليسترول بجرعة ١٪. بعد ٢٨ يوماً، تم جمع عينات الدم من الجرذان للتقدير الكيميائي الحيوي والعينات المأخوذة من دماغ الجرذان ومعالجتها؛ الفحص المجهرى الضوئي باستخدام الهيماتوكليسلين والايوسين فضلاً عن خلايا التمايز ٦٨ هو صبغة كيمياء مناعية لخلايا الدبقية الصغيرة، حيث تم تجانس عينات الدماغ لقياس البيتا أميلويد. زيادة كبيرة في مستوى الدهون في الدم، البيتا أميلويد، أسيتيل كولينستراز، ومستويات مالونديالديهيد، مع