



Molecular study and DNA sequence analysis of *Theileria annulata* in cattle in Al-Hilla, Iraq

A.Q. Jawad  and M.A. Abd Alfatlawi 

Department of Veterinary Microbiology, College of Veterinary Medicine, University of Al-Qadisiyah, Al-Diwaniyah, Iraq

Article information

Article history:

Received August 12, 2022
Accepted November 28, 2022
Available online February 27, 2023

Keywords:

Gene sequencing
Parasitic infection
Theileria annulata
Theileriosis

Correspondence:

M.A. Abd Alfatlawi
monyerr.abd@qu.edu.iq

Abstract

The current work was conducted to unveil the current situation for the infection by *Theileria annulata* in cattle in Al-Hilla City, Iraq. A total of 225 blood samples (200 from suspected infected animals and 25 from clinically healthy animals as a control group) were collected. These samples were subjected to a direct slide-smearing for detection using a microscope and DNA sequencing, targeting the cytochrome b (Cyt b) gene of 10 polymerase chain reaction (PCR) products. The thin smear findings of the 200 suspected cases revealed that 63 (31.5%) were infected with *Theileria* spp., while 115 (57.5%) cases had no *Theileria* but other blood parasites; however, only 22 (11%) suspected cases showed no presence of any parasites. Unsurprisingly, the 25 blood samples from the control group demonstrated no presence of any blood parasite. Moreover, the DNA sequencing demonstrated that the *Theileria* spp. belonged to *T. annulata* species, and these sequences were nucleotide-based similar to Gene-Bank isolates from Tunisia (ON035604, ON035605, ON035606, ON035607, ON035608, ON035609, ON035610, ON035611, ON035612, and ON035613). The present study outcomes indicate that theileriosis is the dominant parasitic infection in cattle in Al-Hilla City and is highly caused by *Theileria annulata*.

DOI: [10.33899/ijvs.2022.135154.2450](https://doi.org/10.33899/ijvs.2022.135154.2450), ©Authors, 2023, College of Veterinary Medicine, University of Mosul.
This is an open access article under the CC BY 4.0 license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction

Only a few of the difficulties faced by Iraqi livestock and dairy industries: a widespread shortage of knowledge among livestock farmers about consuming food, tick control techniques, artificial insemination, and financial damages (1). Parasitism is a major cause of health issues in these farm animals, and most of them are infested with ticks that are considered the natural vector for the transmission of blood parasites. Ticks thrive in Iraq, making it an ideal place to grow and reproduce (2-8). Ticks related to the genera *Hyalomma*, *Rhipicephalus*, and *Ixodes* affect various animals from domestic and wild origins, producing different tick-borne illnesses. In addition to harming the health and production of cattle, theileriosis also costs livestock owners a wide range of financial resources (9-12). Cattle theileriosis can occur due to *Theileria annulata*, an intracellular

protozoan. Several Ixodid tick genera, including *Rhipicephalus*, *Hyalomma*, and *Amblyomma*, are frequently reported as the main vectors for transmitting *Theileria* spp. (13-16). Host bovines undergo the sporogony and merogony phases, whereas ticks develop zygotes and kinetes. When a tick feeds on a host, the parasite enters the host and quickly invades its leukocytes. Once liberated from the parasitized leukocytes, merozoites invade erythrocytes, where they grow into piroplasms (17-22). Conjunctival petechial hemorrhage, swollen lymph nodes, and anemia are symptoms of theileriosis, in addition to high fever, restricted appetite, loss of body weight, and general weakness (23,24). *Theileria* piroplasms are often seen in animals and acute signs and serve as reservoirs for the parasite community (25). It is thus essential to identify carrier animals in epidemiological investigations to determine the level of disease risk and evaluate control measures (26).

The molecular methods used to identify *Theileria annulata* in current work are the main aim of it in cattle in Al-Hilla, Iraq.

Materials and methods

Ethical approve

The study was approved and carried out at the College of Veterinary Medicine, University of Al-Qadisiyah with approval number (P.G, No. 1890 in 2020) during the period September 2021 to February 2022 according to the international guidelines for the care and use of animals.

Blood collection

This study was conducted between September, 2021 to February, 2022. A total of 225 jugular-vein blood-samples (200 from suspected infected animals and 25 from clinically healthy animals as a control group) were collected. The animals were of different ages, from six months to 9 years old, and of both sexes. Blood samples (2 ml/each) were inserted in sterile EDTA treated tubes and transported immediately in an icepack to the Parasitology Laboratory, College of Veterinary Medicine, University of Al-Qadisiyah. Thin blood smears were prepared to identify *Theileria* spp., Then the remaining blood was placed in a deep freezer under -20°C for DNA extraction.

Microscopic examination

Each blood sample was methanol-fixed, Giemsa-stained, and examined using a microscope (100X). The existence of only one piroplasm was reported as positive (27).

DNA Extraction

This extraction of the parasite DNA was made according to the genomic DNA purification Kit supplemented by Geneaid (Korea) and was done depending on the instruction accompanying the kit. As an initial step, 200µl of frozen blood was used as a startup material for the DNA extraction. Ultimately, the final DNA product was Nano Drop - estimated identify its quality and quantity.

PCR

The *Theileria* spp. Was identified using the rRNA gene as a molecular target (primers: F: GAG ACA AGG AAT ATT CTG AGT CC and R: TTA AG TGG CAT ATA ATG ACT TAA GC, (28)). The Cyt b gene was used to identify *Theileria annulata* via sequencing using the primers F: CAG GGC TTT AAC CTA CAA ATT AAC and R: CCC CTC CAC TAA GCG TCT TTC GAC AC, (29), as a molecular target, specifically designed for the current investigation. The 20µl-reaction mixture for the PCR contained 10µl green master mix, 1µl for each upstream primer and downstream primer, 2µl DNA template, 5.5µl for-molecular-use-water, and 0.5 µl MgCl₂. The thermocycler conditions were 95°C for 5mins, (95°C for 35s, 57°C for 35s, and 72°C for the 40s),

and 72°C for 5mins, for the one-cycle for initial denaturation, 39-cycle for (main denaturation, annealing and main extension), and one-cycle for a final extension. For the electrophoresis, 2% agarose gel mixed with 0.5µg/ml ethidium bromide was employed. The bands were then examined utilizing a UV-imager.

Amplicon sequencing analysis

DNA sequencing was conducted for 10 positive-PCR local isolates of *Theileria annulata* from cattle. The PCR products for the Cyt b gene were sent to Macrogen Company in Korea employing the AB DNA sequencing system. The phylogenetic tree analysis was built using MEGA X and the multiple sequence alignment analysis based on Clustal W alignment analysis, and the related evolutionary distances were calculated employing the maximum composite likelihood method via the phylogenetic tree UPGMA method. Comparisons were made using the sequences of the local isolates against isolates from the NCBI-Blast. Finally, the sequences of the local isolates were deposited into the NCBI GenBank get accession numbers.

Results

Microscopic examination

The thin blood smears findings of the 200 suspected cases revealed that 63 (31.5%) were infected with *Theileria* spp., while 115 (57.5%) cases had no *Theileria* but other blood parasites; however, only 22 (11%) suspected cases showed no presence of any parasites. Unsurprisingly, the 25 blood samples from the control group demonstrated no presence of any blood parasite.

DNA sequencing

The DNA sequencing demonstrated that the *Theileria* spp. belonged to *T. annulata* species, and these sequences were nucleotide-based similar to Gene-Bank isolates from Tunisia (ON035604, ON035605, ON035606, ON035607, ON035608, ON035609, ON035610, ON035611, ON035612, and ON035613). (Figure 1).

Discussion

Dairy sector expansion has been hindered by tick-borne diseases (TBDs) that generate significant economic consequences. According to earlier investigations, *T. annulata* was detected in 33 and 24% of cattle in Pakistan from two districts. Also, in Pakistan, the occurrence of *T. annulata* in cattle in different areas was revealed to be 33, 30, 28, 23.7, 21, 19, and 18.8% (30-35). Additionally, *T. annulata* infection in cattle has been documented in many nations that fall within the tropical or subtropical climate zones. *T. annulata* prevalence in cattle was 23.3, 20, 25.4, 18.2, and 1.9% in India, Egypt, Algeria, Northwest China, and Saudi Arabia, respectively. These data from the

countries mentioned above regarding the infection rates lower than the rate of the current study, probably, due to some failure in the control programs of ticks in the current study areas. Differences in tick eradication strategies, environment compatibility, farm control, husbandry techniques, and abiotic conditions at sampling locations might cause differences and help in the varying infection rates of *T. annulata* from one site to another (30,36,37).

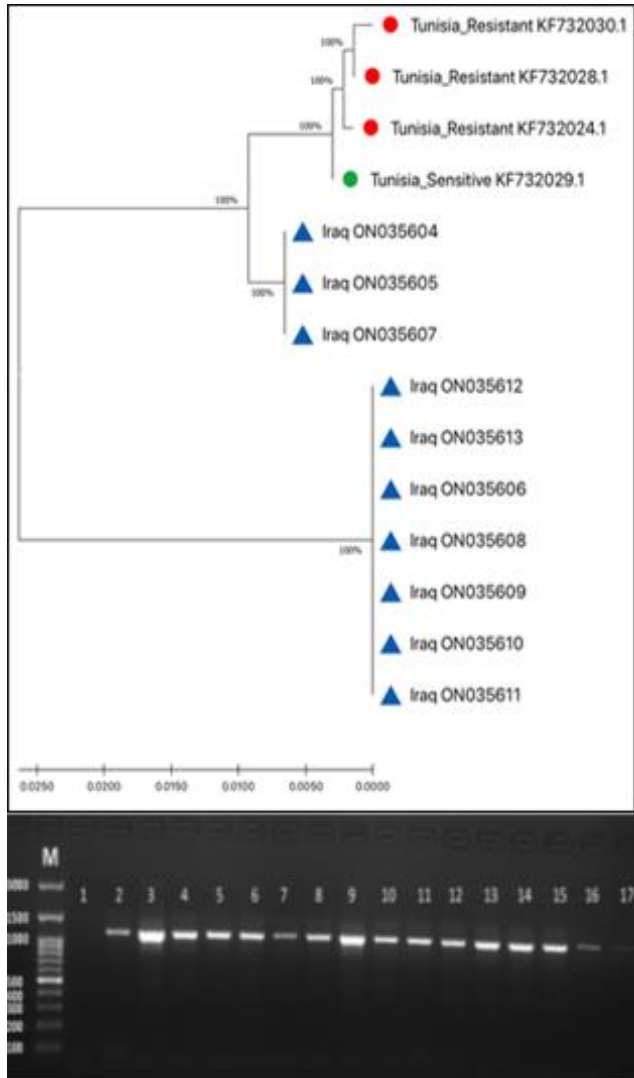


Figure 1: *Theileria annulata* based phylogenetic tree (Cyt b gene) of the study detected sequences (blue triangles + Accession numbers).

The primary ingredient for the evolution of microorganisms is genetic diversity (37), so the genetic variety of *T. annulata* in a host animal enables the parasite to avoid the host's immune system. Chromosomal recombination in tick vectors throughout sexual reproduction is how *T. annulata* acquires its genetic variety

(38). Factors like genetic drift and mutation strengthen their genetic variation. Because of this, creating control methods (such as vaccinations and pharmacological treatments) depends on parasite populations acquiring genetic diversity (39). The foundation for genetic differences and evolutionary links between species may be found through phylogenetic analysis. The piroplasm population has recently been studied using molecular markers, including 18S rRNA, ITS1, ITS2, and the Cyt b gene, to identify genetic associations between local and global isolates (40). Marker genes are essential tools for detecting the evolutionary connection between species because of the occurrence of both highly conserved and changeable areas of the genome (41,42). There are many *T. annulata* genetic diversity data from Iraq, especially Al-Hilla City. The current similarity between the present study isolates and the GeneBank isolates could be due to importing cattle infested with ticks from different countries to Iraq, such as India. It could be due to the travel of the tick vectors from different countries to Iraq via some tools, including migrating birds, in which new species of *T. annulata* might be brought in, and new genetic differentiation might occur in Iraq.

Conclusion

The present work demonstrates that cattle from Al-Hilla City, Iraq, were highly infected with *Theileria annulata* compared to those from other countries. The current study shows links between the current identified local and some global isolates of the protozoan.

Acknowledgments

The authors thank Professor Jabbar Ahmed Alssady, Dean of College of Veterinary Medicine, University of Al-Qadisiyah, Iraq, for technical assistance.

Conflict of interests

The authors have not received any funding or benefits from industry, agency of financing, or elsewhere to conduct this study.

References

1. Jabbar A, Abbas T, Sandhu ZD, Saddiqi HA, Qamar MF, Gasser RB. Tick-borne diseases of bovines in Pakistan: Major scope for future research and improved control. *Parasit Vectors*. 2015;8(1):283-95. DOI: [10.1186/s13071-015-0894-2](https://doi.org/10.1186/s13071-015-0894-2)
2. Hassan MA, Liu J, Rashid M, Iqbal N, Guan G, Yin H, Iqbal N. Molecular survey of piroplasm species from selected areas of China and Pakistan. *Parasit Vectors*. 2018;11(1):457-63. DOI: [10.1186/s13071-018-3035-x](https://doi.org/10.1186/s13071-018-3035-x)
3. Alfatlawi MA, Jasim AA, Jarad NE, Khlaif SF. Clinical and molecular identification of ruling *Theileria annulata* strains in cattle calves in Al-Diwaniyah province, Iraq. *Iraqi J Vet Sci*. 2021;35(1):115-119. DOI: [10.33899/ijvs.2020.126429.1319](https://doi.org/10.33899/ijvs.2020.126429.1319)

4. El-Seify MA, Helmy NM, Elhawary NM, Sorour SS, Soliman AM. Use molecular techniques as an alternative tool for diagnosis and characterization of *Theileria equi*. Iraqi J Vet Sci. 2018;32(1):5-11. DOI: [10.33899/ijvs.2018.153787](https://doi.org/10.33899/ijvs.2018.153787)
5. Weir W, Karageç T, Gharbi M, Simuunza M, Aypak S, Aysul N. Population diversity and multiplicity of infection in *Theileria annulata*. Int J Parasitol. 2011;41(2):193-203. DOI: [10.1016/j.ijpara.2010.08.004](https://doi.org/10.1016/j.ijpara.2010.08.004)
6. Al-Fatlawi MA, Ali MJ, Al-Bayati HH. Morphological and phylogenetic study of *Hyalomma anatolicum* in Al-Najaf, Iraq. Iraqi J Vet Sci. 2018;32(2):261-6. DOI: [10.33899/ijvs.2019.153860](https://doi.org/10.33899/ijvs.2019.153860)
7. Oryan A, Namazi F, Sharifiyazdi H, Razavi M, Shahriari R. Clinicopathological findings of a natural outbreak of *Theileria annulata* in cattle: An emerging disease in southern Iran. Parasitol Res. 2013;112(1):123-7. DOI: [10.1007/s00436-012-3114-4](https://doi.org/10.1007/s00436-012-3114-4)
8. Ali MJ, Atiyah WR, Al-Fatlawi MA, Khlaif SF. Genotypic analysis of ticks species infesting cattle in Al-Diwaniyah abattoir. Iraqi J Vet Sci. 2021;35(4):673-677. DOI: [10.33899/ijvs.2020.127772.1525](https://doi.org/10.33899/ijvs.2020.127772.1525)
9. Parveen A, Ashraf S, Khan A, Asif M, Iqbal F. Tick and tick-borne diseases in Pakistan. 1st ed. USA: Nova Science; 2021. 49-80 p.
10. Gharaya G, Rakha NK, Maan S, Kumar A, Kumar T, Jhambh R. Comparative evaluation of polymerase chain reaction assay with microscopy for detection of asymptomatic carrier state of theileriosis in a herd of crossbred cattle. Vet World. 2016;9(9):1039-42. DOI: [10.14202/vetworld.2016.1039-1042](https://doi.org/10.14202/vetworld.2016.1039-1042)
11. Kundave VR, Patel AK, Patel PV, Hasnani JJ, Joshi CG. Detection of theileriosis in cattle and buffaloes by polymerase chain reaction. J Parasit Dis. 2015;39(3):508-513. DOI: [10.1007/s12639-013-0386-2](https://doi.org/10.1007/s12639-013-0386-2)
12. Ayadi O, Gharbi M, Benchikh MC. Haematological and biochemical indicators of tropical theileriosis diseased cattle in Wilaya of Sétif (north east Algeria). J Parasit Dis. 2017;41(2):538-42. DOI: [10.1007/s12639-016-0846-6](https://doi.org/10.1007/s12639-016-0846-6)
13. Mans BJ, Pienaar R, Latif AA. A review of *Theileria* diagnostics and epidemiology. Int J Parasitol Parasit Wildl. 2015;4(1):104-18. DOI: [10.1016/j.ijppaw.2014.12.006](https://doi.org/10.1016/j.ijppaw.2014.12.006)
14. Silatsa BA, Simo G, Githaka F, Kamga R, Oumarou F, Tiambo CK, Machuka E, Domelevo J, Odongo D, Bishop R, Kuate J, Njikou F, Djikeng A, Pelle R. First detection of *Theileria parva* in cattle from Cameroon in the absence of the main tick vector *Rhipicephalus appendiculatus*. Trans Emerg Dis. 2020;67(1):68- 78. DOI: [10.1111/tbed.13425](https://doi.org/10.1111/tbed.13425)
15. Al-Hosary AT, Nordengrahen N. New approach to use blood smears for diagnosis of bovine theileriosis. Indian J Anim Res. 2018;10(1):1-4. DOI: [10.18805/ijar.B-870](https://doi.org/10.18805/ijar.B-870)
16. Sharifiyazdi H, Namazi F, Oryan A, Shahriari R, Razavi M. Point mutations in the *Theileria annulata* cytochrome b gene is associated with buparvaquone treatment failure. Vet Parasitol. 2012;187(3-4):431-5. DOI: [10.1016/j.vetpar.2012.01.016](https://doi.org/10.1016/j.vetpar.2012.01.016)
17. Sivakumar T, Hayashida K, Sugimoto C, Yokoyama N. Evolution and genetic diversity of *Theileria*. Infect Genet Evol. 2014;27(10):250-63. DOI: [10.1016/j.meegid.2014.07.013](https://doi.org/10.1016/j.meegid.2014.07.013)
18. Liu J, Guan G, Yin H. *Theileria annulata*. Trends Parasitol. 2022;38(3):265-6. DOI: [10.1016/j.pt.2021.11.001](https://doi.org/10.1016/j.pt.2021.11.001)
19. Chatanga E, Mosssad E, Abdo AH, Amin AS, Katakura K, Nakao R. Evidence of multiple point mutations in *Theileria annulata* cytochrome b gene incriminated in buparvaquone treatment failure. Acta Trop. 2019;191:128-32. DOI: [10.1016/j.actatropica.2018.12.041](https://doi.org/10.1016/j.actatropica.2018.12.041)
20. Alsaad K, Sulieman EG, Al-Obaidi QT. Theileriosis in newborn calves in Mosul, Iraq. Bas J Vet Res. 2013;12(1):265-274. DOI: [10.33762/bvetr.2013.76207](https://doi.org/10.33762/bvetr.2013.76207)
21. Dandasena D, Bhandari V, Sreenivasamurthy GS, Murthy S, Roy S, Bhanot V. A real-time PCR based assay for determining parasite to host ratio and parasitaemia in the clinical samples of bovine theileriosis. Sci Rep. 2018;8(1):1-7. DOI: [10.1038/s41598-018-33721-3](https://doi.org/10.1038/s41598-018-33721-3)
22. Shinuo CO, Zhang S, JIA L, Xue S, Yu L, Kamyngkird K, Moumouni PA, Moussa AE, Zhou M, Zhang Y. Molecular Detection of *Theileria* Species in sheep from northern China. J of Vet Med Sci. 2013;75(9):1227-1230. DOI: [10.1292/jvms.13-0028](https://doi.org/10.1292/jvms.13-0028)
23. Klaiif SF, Abid AJ, Al-Fatlawi MA, Ali MJ. Major-surface-protein-4-gene-based detection of *Anaplasma marginale* isolated from sheep in Al-Diwaniyah province, Iraq. Iraqi J Vet Sci. 2022;36(1):85-88. DOI: [10.33899/ijvs.2021.129230.1635](https://doi.org/10.33899/ijvs.2021.129230.1635)
24. Watts JG, Playford MC, Hickey KL. *Theileria orientalis*: A review. NZ Vet J. 2016;64(1):3-9. DOI: [10.1080/00480169.2015.1064792](https://doi.org/10.1080/00480169.2015.1064792)
25. Abid K, Bukhari S, Asif M, Sattar A, Arshad M, Aktas M, Özübek S, Shaik RS, Iqbal F. Molecular detection and prevalence of *Theileria ovis* and *Anaplasma marginale* in sheep blood samples collected from Layyah district in Punjab, Pakistan. Trop Anim Heal Prod. 2021;53(4):1-9. DOI: [10.1007/s11250-021-02870-5](https://doi.org/10.1007/s11250-021-02870-5)
26. Santos M, Soares R, Costa P, Amaro A, Inácio J, Gomes J. Revisiting the Tams1-encoding gene as a species-specific target for the molecular detection of *Theileria annulata* in bovine blood samples. Ticks Tick Borne Dis. 2013;4(1-2):72-7. DOI: [10.1016/j.ttbdis.2012.07.006](https://doi.org/10.1016/j.ttbdis.2012.07.006)
27. Bahrami S, Tabandeh MR, Nikbin A, Alborzi AR, Ghaadrnan AR. Prevalence and phylogenetic analysis of *Theileria equi* in Iranian dromedaries. Arch Razi Inst. 2016;71(3):169-75. DOI: [10.22034/ari.2016.106970](https://doi.org/10.22034/ari.2016.106970)
28. Habibi G, Sepahvand-Mohammadi E, Afshari A, Bozorgi S. Molecular detection of *Theileria* spp. and *Babesia ovis* infection in sheep in Baneh, Iran. Arch Razi Inst. 2020;75(2):289-96. DOI: [10.22092/ari.2019.125136.1297](https://doi.org/10.22092/ari.2019.125136.1297)
29. Mhadhbi M, Chaouch M, Ajroud K, Darghouth MA, BenAbderrazak S. Sequence polymorphism of cytochrome b gene in *Theileria annulata* Tunisian isolates and its association with buparvaquone treatment failure. PLoS One. 2015;10(6):e0129678-88. DOI: [10.1371/journal.pone.0129678](https://doi.org/10.1371/journal.pone.0129678)
30. Parveen A, Alkhaibari AM, Asif M, Almohammed HI, Naqvi Z, Khan A, Aktas M, Ozubek S, Farooq M, Iqbal F. Molecular epidemiology of *Theileria annulata* in cattle from two districts in Punjab (Pakistan). Anim Open. 2021;11(12):3443-54. DOI: [10.3390/ani11123443](https://doi.org/10.3390/ani11123443)
31. Farooqi SH, Ijaz M, Saleem MH, Rashid MI, Ahmad SS, Islam S. Prevalence and molecular diagnosis of *Theileria annulata* in bovine from three distinct zones of Khyber Pakhtunkhwa province, Pakistan. J Anim Plant Sci. 2017;27(6):1836-41. [available at]
32. Zeb J, Shams S, Din IU, Ayaz S, Khan A, Nasreen N, Khan S, Khan MS, Senbill H. Molecular epidemiology and associated risk factors of *Anaplasma marginale* and *Theileria annulata* in cattle from north-western Pakistan. Vet Parasitol. 2020;279(2):109049. DOI: [10.1016/j.vetpar.2020.109044](https://doi.org/10.1016/j.vetpar.2020.109044)
33. Khattak RM, Rabib M, Khan Z, Ishaq M, Hameed H, Taqddus A, Faryal M, Durrani S, Gillani A, Allahyar R, Shaikh RS, Khan MA, Ali M, Iqbal F. A comparison of two different techniques for the detection of a blood parasite, *Theileria annulata*, in cattle from two districts in Khyber Pukhtoon Khwa province (Pakistan). Parasite. 2012;19(1):91-5. DOI: [10.1051/parasite/2012191091](https://doi.org/10.1051/parasite/2012191091)
34. Parveen A, Ashraf S, Aktas M, Ozubek S, Iqbal F. Molecular epidemiology of *Theileria annulata* infection of cattle in Layyah district, Pakistan. Exp Appl Acarol. 2021;83(3):461-73. DOI: [10.1007/s10493-021-00595-6](https://doi.org/10.1007/s10493-021-00595-6)
35. Shahnawaz S, Ali M, Aslam MA, Fatima R, Chaudhry ZI, Hassan MU, Iqbal F. A study on the prevalence of a tick-transmitted pathogen, *Theileria annulata*, and hematological profile of cattle from southern Punjab (Pakistan). Parasitol Res. 2011;109(4):1155-60. DOI: [10.1007/s00436-011-2360-1](https://doi.org/10.1007/s00436-011-2360-1)
36. Alanazi AD, Alouffi AS, Alshahrani MY, Alyousif MS, Abdullah HM, Allam AM, Elsayy B, Abdel-Shafy S, Alsolami S, Khan A, Iqbal F. A report on tick burden and molecular detection of tick-borne pathogens in cattle blood samples collected from four regions in Saudi Arabia. Ticks Tick Borne Dis. 2021;12(3):101652-10172. DOI: [10.1016/j.ttbdis.2021.101652](https://doi.org/10.1016/j.ttbdis.2021.101652)
37. Guo H, Yin C, Galon EM, Du J, Gao Y, Adjou Moumouni PF, Liu M, Efstratiou A, Lee SH, Li J, Ringo AE, Wang G, Li Y, Tumwebaze MA, Xuan X. Molecular survey and characterization of *Theileria annulata* and *Ehrlichia ruminantium* in cattle from northwest China. Parasitol Int. 2018;67(6):679-83. DOI: [10.1016/j.parint.2018.06.011](https://doi.org/10.1016/j.parint.2018.06.011)
38. Roy S, Bhandari V, Barman M, Kumar P, Bhanot V, Arora JS, Singh S, Sharma P. Population genetic analysis of the *Theileria annulata* parasites identified limited diversity and multiplicity of infection in the

- vaccine from India. Front Microbiol. 2021;11(1):579929-39. DOI: [10.3389/fmicb.2020.579929](https://doi.org/10.3389/fmicb.2020.579929)
39. Al-Hamidhi S, Tageldin MH, Weir W, Al-Fahdi A, Johnson EH, Bobade P, Al-Qamashou B, Pereira AP, Thompson JI, Kinnaird J, Sheils B, Tait A, Babiker HA. Genetic diversity and population structure of *Theileria annulata* in Oman. PLoS One. 2015;10(10):e0139581-96. DOI: [10.1371/journal.pone.0139581](https://doi.org/10.1371/journal.pone.0139581)
40. Habibi G. Phylogenetic Analysis of *Theileria annulata* infected cell line S15 Iran vaccine strain. Iran J Parasitol. 2012;7(2):73-81. [\[available at\]](#)
41. Gupta RS. Impact of genomics on the understanding of microbial evolution and classification: the importance of Darwin's views on classification. FEMS Microbiol Rev. 2016;40(4):520-53. DOI: [10.1093/femsre/fuw011](https://doi.org/10.1093/femsre/fuw011)
42. Kaaboub EA, Ouchene N, Ouchene NA, Dahmani A, Ouchtati I, Haif A, Khelef D. Investigation of the principal vectors of abortive diseases in one-humped camels (*Camelus dromedarius*). Iraqi J Vet Sci. 2021;35(3):411-415. DOI: [10.33899/ijvs.2020.126914.1415](https://doi.org/10.33899/ijvs.2020.126914.1415)

دراسة جزيئية والتحليل الجيني لطفيلي الثلاريا انيولاتا في الأبقار في مدينة الحلة، العراق

آيات قاسم جواد و منير عبد الأمير عبد الفتلاوي

فرع الأحياء المجهرية البيطرية، كلية الطب البيطري، جامعة القادسية، الديوانية، العراق

الخلاصة

تم تنفيذ العمل الحالي لكشف النقاب عن الوضع الحالي لعدوى طفيلي الثلاريا انيولاتا في الماشية في مدينة الحلة، العراق. في هذه الدراسة تم جمع إجمالي ٢٢٥ عينة دم (٢٠٠ من الحيوانات المشتبه بإصابتها و ٢٥ عينة من الحيوانات السليمة سريريا). تم إخضاع هذه العينات إلى الفحص المجهرى وفحص تسلسل الحمض النووي، باستهداف جين السيتوكروم ب، لعشرة منتجات من تفاعل أنزيم البلمرة المتسلسل. أظهرت نتائج المسحة الرقيقة للحالات المشتبه بها البالغ عددها ٢٠٠ حالة إصابة ٦٣ (٣١,٥٪) بالثلبيريا، بينما كانت ١١٥ حالة (٥٧,٥٪) مصابة بأنواع أخرى من العدوى الطفيلية في الدم. ومع ذلك، لم تظهر سوى ٢٢ حالة (١١٪) مشتبه فيها عدم وجود أي طفيليات. بالإضافة إلى ذلك، لم تظهر أي عدوى طفيلية في ٢٥ عينة دم. علاوة على ذلك، أظهر تسلسل الحمض النووي أن أنواع الثاليريا تنتمي إلى أنواع الانيولاتا، وكانت هذه التسلسلات تعتمد على تشابه القواعد النيتروجينية بين العزلات المحلية للدراسة الحالية وعزلات بنك الجينات من تونس (ON035604, ON035605, ON035606, ON035607, ON035608, ON035609, ON035610, ON035611, ON035612, and ON035613). تشير نتائج الدراسة الحالية إلى أن مرض الثاليريا هي العدوى الطفيلية السائدة في الماشية في مدينة الحلة وتسببها بشكل كبير نوع الثلاريا انيولاتا.